

RESEARCH ARTICLE

The pyrostatins A and B do not inhibit N-acetyl- β -D-glucosaminidase

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Abstract

The compounds pyrostatin A and B, isolated from *Streptomyces sp.* SA-3501 have been reported as N-acetyl- β -D-glucosaminidase inhibitors with inhibition constants in the micromolar range. Recently, a comparison of NMR spectral data of the pyrostatins has led to a structural revision of the pyrostatins. It was shown that the pyrostatins A and B are identical to the ectoines 5-hydroxyectoine and ectoine, respectively. Ectoines are known as compatible osmolytes in many halophilic and stress-tolerant bacteria. We have performed enzymatic experiments demonstrating that neither ectoine nor 5-hydroxyectoine exhibit an inhibitory effect on N-acetyl- β -D-glucosaminidase. The previously reported inhibition of N-acetyl- β -D-glucosaminidase by pyrostatins A and B may thus be due to the contamination of the compound preparations with a strong N-acetyl- β -D-glucosaminidase inhibitor with an inhibition constant (K_i) in the nanomolar range, as has been reported in other *Streptomyces* species.

Keywords: ectoine; hydroxyectoine; pyrostatin A; pyrostatin B; N-acetyl- β -D-glucosaminidase

Abbreviation: GlcNAc-ase: N-acetyl- β -D-glucosaminidase

Introduction

N-acetyl-beta-D-glucosaminidase (GlcNAc-ase; E.C. 3.2.1.30) catalyzes the hydrolytic release of N-acetyl-beta-D-glucosamine from glycoproteins. Previously, Aoyama et al. (1995) purified and determined two GlcNAc-ase inhibitors, pyrostatin A and B, from *Streptomyces sp.* SA-3501, which was isolated from a marine environment [1]. The structures of pyrostatin A and B were determined by ¹H and ¹³C-NMR analysis to be 4-hydroxy-2-amino-1-methylpyrrolidine-5-carboxylic acid and 2-imino-1-methylpyrrolidine-5-carboxylic acid respectively. It was reported that both pyrostatin A and B were competitive inhibitors on GlcNAc-ase with inhibition constants (K_i) of 1.7×10^{-6} M and 2.0×10^{-6} M respectively [1].

As part of a search for biologically active secondary metabolites, Castellanos et al. 2006 discovered 2-imino-1-methylpyrrolidine-5-carboxylic acid in the organic extracts of the sponge *Cliona tenius* [2]. The structure was determined using spectral analysis, however the data did not match those published for pyrostatin B by Aoyama et al. To clarify this discrepancy Castellanos et al. performed the total synthesis of 2-imino-1-methylpyrrolidine-5-carboxylic acid. The analysis of the synthesized compound confirmed their

proposed structure and thus showed that the structure proposed by Aoyama et al. for pyrostatin B is incorrect.

Based on a comparison with the NMR data published in [1], Castellanos et al. found that the data reported for pyrostatin A and pyrostatin B match those of 5-hydroxyectoine and ectoine respectively.

Ectoine and hydroxyectoine (Figure 1) are known as compatible solutes and are widespread in halotolerant and halophilic bacteria. Several species of the genus *Streptomyces*, from which the pyrostatins have been isolated, are known to produce ectoine and hydroxyectoine [3–8]. Compatible solutes like the ectoines can be accumulated by microorganisms up to very high concentrations and are generally known as chemically inert. Consequently, no enzyme-inhibitory effect has been described for these compounds so far. We therefore re-examined whether pyrostatin A (5-hydroxyectoine) and pyrostatin B (ectoine) inhibit GlcNAc-ase, as described by Aoyama et al. [1].

The enzymatic activity of GlcNAc-ase from bovine kidney in the presence and absence of various ectoine concentrations was measured. 2-acetamido-2-deoxy-D-glucono-1,5-lactone, a known strong inhibitor of GlcNAc-ase was used as a positive control for inhibition [9].

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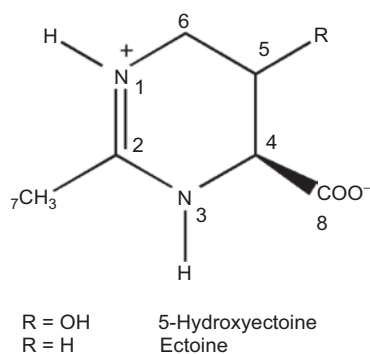


Figure 1. Structure of ectoine and 5-hydroxyectoine.

Material and methods

Materials

GlcNAc-ase and 4-nitrophenyl N-acetyl-β-D-glucosaminide were purchased from Sigma-Aldrich (Seelze, Germany). 2-Acetamido-2-deoxy-D-glucono-1,5-lactone was obtained from Chemos GmbH (Regenstauf, Germany). Ectoine and 5-hydroxyectoine (isolated from *Halomonas elongata*) were from bitop AG (Witten, Germany). All other chemicals were of standard analytical grade from Merck KGaA (Darmstadt, Germany).

Methods

The enzyme activity of GlcNAc-ase was determined by measuring the amount of liberated *p*-nitrophenol when using 4-nitrophenyl N-acetyl-β-D-glucosaminide as a substrate [10]. The reaction mixture contained 790 μL 0.1 M sodium citrate buffer (pH 5.0), 100 μL 4-nitrophenyl N-acetyl-β-D-glucosaminide in final concentrations ranging from 0.3 mM to 2.5 mM. Ectoine, 5-hydroxyectoine and 2-acetamido-2-deoxy-D-glucono-1,5-lactone were added in a volume of 10 μL to the assay mixture, whereas water was used for reference measurements. After addition of 100 μL GlcNAc-ase (0.17 U/mL) the mixture was incubated in 1.5 mL semi-micro cuvettes. After 10 min of incubation the reaction was stopped by adding 10 μL 1 M glycine-sodium hydroxide buffer (pH 12) to the reaction mixture. The absorbance at 400 nm was measured using a Pharmacia Ultrospec 3000 Photometer.

Results and discussion

2-acetamido-2-deoxy-D-glucono-1,5-lactone shows competitive inhibition of GlcNAc-ase with an inhibition constant (K_i) of 0.036 μM at pH 4.25 [9]. It was therefore chosen as a positive control to clarify if ectoine and 5-hydroxyectoine also show an inhibitory effect on GlcNAc-ase. For the inhibition of GlcNAc-ase two different concentrations (0.11 μM and 1.1 μM) of 2-acetamido-2-deoxy-D-glucono-1,5-lactone were used. As indicated in Figure 2, 2-acetamido-2-deoxy-D-glucono-1,5-lactone shows typical characteristics of a competitive inhibitor, with a calculated K_i value of 0.18 μM at pH 5.0.

To measure the inhibition constants of ectoine and 5-hydroxyectoine on GlcNAc-ase, we used several concentrations

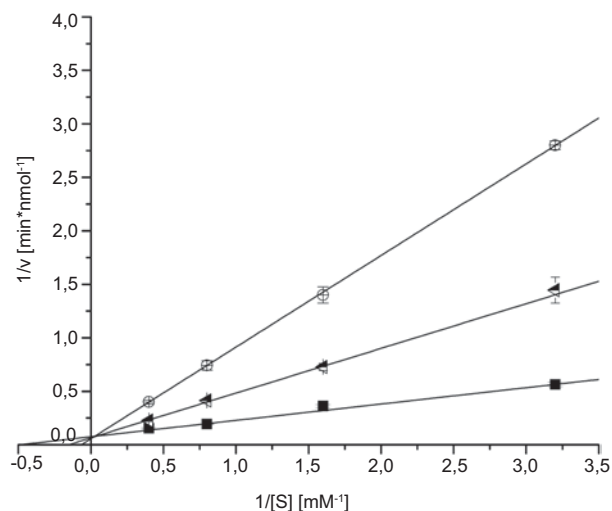


Figure 2. Lineweaver-Burk plots for inhibition of N-acetyl-β-D-glucosaminidase by 2-acetamido-2-deoxy-D-glucono-1,5-lactone at different concentrations; 0 μM (■), 0.11 μM (◄), 1.1 μM (⊕). Each data point shows the mean value of five independent measurements.

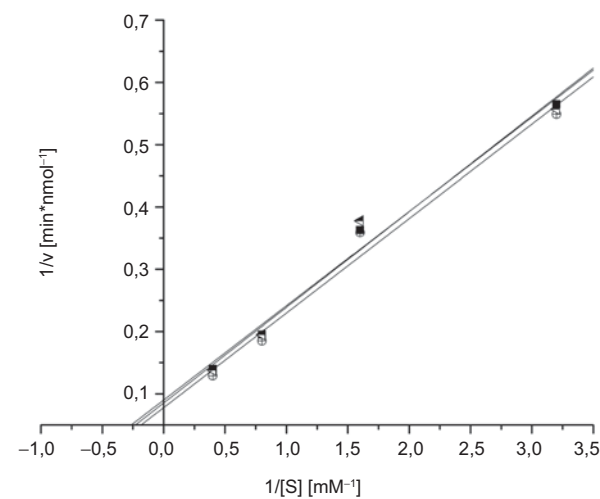


Figure 3. Lineweaver-Burk plots for inhibition of N-acetyl-β-D-glucosaminidase by ectoine and 5-hydroxyectoine; blank (■), 210 μM ectoine (◄), 210 μM 5-hydroxyectoine (⊕). Each data point shows the mean value of five independent measurements.

in the range of 0.2 μM to 210 μM of ectoine and 5-hydroxyectoine. As shown in Figure 3, even the highest concentration used does not induce an inhibitory effect of ectoine or 5-hydroxyectoine.

Comparison of the ^1H and ^{13}C -NMR spectral data for pyrostatin B [1], and ectoine [11] as shown in Tables 1 and 2 shows virtually no difference in either the proton or carbon chemical shifts.

In contrast comparison of the ^1H and ^{13}C -NMR data for the synthetic 2-imino-1-methylpyrrolidone-5-carboxylic acid [2] does not match those reported for pyrostatin B (data not shown).

In summary it can be concluded that the substance pyrostatin B described by Aoyama et al. is in fact ectoine,

Table 1. Comparison of ¹H-NMR data of ectoine [11] and pyrostatin B [1]

δ/ppm	δ/ppm				^x J _{y,z}	^x J _{y,z}
Ectoine	Pyrostatin B	Mult.	Proton	Int.	Ectoine	Pyrostatin B
2.11	2.13	m	5	2H	-	-
2.22	2.24	s	7	3H	-	-
3.28	3.30	ddd	6	1H	13.5; 8.4; 4.8	14.0; 8.6; 5.0
3.44	3.46	ddd	6	1H	13.5; 5.4; 5.4	14.0; 5.6; 5.6
4.06	4.07	dd	4	1H	5.4; 5.4	5.6; 5.6

Table 2. Comparison of ¹³C-NMR data of ectoine [11] and pyrostatin B [1]

δ/ppm Ectoine (Lit)	δ/ppm Pyrostatin B (Lit)	Mult.	Carbon
19.1	18.9	CH ₃	7
23.6	22.1	CH ₂	5
38.0	38.0	CH ₂	6
53.6	53.9	CH	4
161.5	161.2	Cq	2
177.0	177.4	Cq	8

and furthermore pyrostatin A is 5-hydroxyectoine, as proposed by Castellanos et al. (2006). Furthermore we demonstrated that these compounds do not have an inhibitory effect towards GlcNAc-ase. One possible explanation for the inhibition observed by Aoyama et al. is a contamination of the pyrostatin preparations with a highly potent GlcNAc-ase inhibitor, possibly similar to the *Streptomyces* compound nagstatin, with a K_i for GlcNAc-ase in the nanomolar range [12].

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Declaration of interest: The authors report no conflicts of interest. The authors alone are responsible for the content and writing of the paper.

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